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| **Food and Environmental Proficiency Testing Unit**  Store LENTICULE disc samples @ -20°C± 5ºC  **Laboratory Identification no. Lab No.** Dispatch date: **XX XXXX 20XX** Final date for return of results: **XX XXXX 20XX** | | | | |
| **Contact details:**  The Organisers - FEPTU  Public Health England  61 Colindale Avenue,  London, NW9 5EQ, UK.  Fax: +44 (0) 20 8200 8264  Tel: +44 (0) 20 8327 7119  e-mail: foodeqa@phe.gov.uk | | **RESULTS TO BE RETURNED VIA ONLINE SURVEY AS PER EMAIL SENT TO YOU** | | |
| **0006** | | | | |
|  | | | | |
| **Shiga toxin producing *Escherichia coli* (STEC) Request/Report Form** | | | |
| **Distribution No.: STXXX** | | | **Sample numbers**: **STX00X & STX00X** |
| Download the sample Instruction sheet | | | [www.gov.uk/government/publications/shiga-toxin-escherichia-coli-scheme-sample-instruction-sheet](http://www.gov.uk/government/publications/shiga-toxin-escherichia-coli-scheme-sample-instruction-sheet) |
| Download the safety data sheet: | | | [www.gov.uk/government/publications/safety-data-sheet-for-lenticules](http://www.gov.uk/government/publications/safety-data-sheet-for-lenticules) |
| ***If you cannot examine any of these samples return your results as ‘Not examined’*** | | | |
| **Request:** | | | 1. Examine samples for STEC genes |
|  | |  | |

**Food and Environmental Proficiency Testing Unit**

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| **Laboratory Identification no. Lab No.** |  |

**SAMPLE: STX00X**

**STEC / VTEC / EHEC (Shiga / verocytotoxic / enterohemorrhagic) testing:-**

|  | **Target Genes** | **Positive (circle)** | **CT value** |  |  | **Not examined** | **Serogroup**  **(e.g. O157)** | **Positive (circle)** | **CT value** | **Serotype**  **(e.g. H7)** | **Positive (circle)** | **CT value** |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **STEC virulence gene detection** | *stx*  *(stx* 1/2 combined) | Yes / No / NE |  | **STEC identification gene detection** | 🞏 | O157 | Yes / No / NE |  |  | Yes / No / NE |  |
| *stx* 1 | Yes / No / NE |  | O26 | Yes / No / NE |  |  | Yes / No / NE |  |
| *stx* 2 | Yes / No / NE |  | O103 | Yes / No / NE |  |  | Yes / No / NE |  |
| *eae* | Yes / No / NE |  | O104 | Yes / No / NE |  |  | Yes / No / NE |  |
| Other …….. | Yes / No / NE |  | O111 | Yes / No / NE |  |  | Yes / No / NE |  |
|  |  |  | O145 | Yes / No / NE |  |  | Yes / No / NE |  |
|  |  |  | Other………. | Yes / No / NE |  |  | Yes / No / NE |  |

NE = Not Examined

**Interpretation of results:-**

|  |  |  |
| --- | --- | --- |
| **Select interpretation of your results** | | **Explanation** |
| 🞏 | Not examined / Not applicable |  |
| 🞏 | STEC not detected in the test portion of x g or xml |  |
| 🞏 | Presumptive detection of STEC in the test portion of x g or x ml |  |
| 🞏 | Presumptive detection of STEC causing the attaching and effacing lesion in the test portion of x g or x ml |  |
| 🞏 | Presumptive detection of STEC of xx serogroup in the test portion of x g or x ml |  |
| 🞏 | Other: |  |

**Food and Environmental Proficiency Testing Unit**

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| **Laboratory Identification no. Lab No.** |  |

**SAMPLE: STX00X**

**STEC / VTEC / EHEC (Shiga / verocytotoxic / enterohemorrhagic) testing:-**

|  | **Target Genes** | **Positive (circle)** | **CT value** |  |  | **Not examined** | **Serogroup**  **(e.g. O157)** | **Positive (circle)** | **CT value** | **Serotype**  **(e.g. H7)** | **Positive (circle)** | **CT value** |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **STEC virulence gene detection** | *stx*  *(stx* 1/2 combined) | Yes / No / NE |  | **STEC identification gene detection** | 🞏 | O157 | Yes / No / NE |  |  | Yes / No / NE |  |
| *stx* 1 | Yes / No / NE |  | O26 | Yes / No / NE |  |  | Yes / No / NE |  |
| *stx* 2 | Yes / No / NE |  | O103 | Yes / No / NE |  |  | Yes / No / NE |  |
| *eae* | Yes / No / NE |  | O104 | Yes / No / NE |  |  | Yes / No / NE |  |
| Other …….. | Yes / No / NE |  | O111 | Yes / No / NE |  |  | Yes / No / NE |  |
|  |  |  | O145 | Yes / No / NE |  |  | Yes / No / NE |  |
|  |  |  | Other………. | Yes / No / NE |  |  | Yes / No / NE |  |

NE = Not Examined

**Interpretation of results:-**

|  |  |  |
| --- | --- | --- |
| **Select interpretation of your results** | | **Explanation** |
| 🞏 | Not examined / Not applicable |  |
| 🞏 | STEC not detected in the test portion of x g or xml |  |
| 🞏 | Presumptive detection of STEC in the test portion of x g or x ml |  |
| 🞏 | Presumptive detection of STEC causing the attaching and effacing lesion in the test portion of x g or x ml |  |
| 🞏 | Presumptive detection of STEC of xx serogroup in the test portion of x g or x ml |  |
| 🞏 | Other: |  |

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| **Laboratory Identification no. Lab No.** |  |

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| **Any further comments:-** |

**Authorised by: Date reported:**

**Laboratory Identification no. Lab No**

**METHOD QUESTIONNAIRE – not all findings will be presented in the scheme report (include as much detail as possible or references):**

|  |  |  |
| --- | --- | --- |
| **1. Do you exactly follow ISO/TS 13136:2012**  *Microbiology of food and animal feed -- Real-time polymerase chain reaction (PCR)-based method for the detection of food-borne pathogens - Horizontal method for the detection of Shiga toxin-producing* Escherichia coli *(STEC) and the determination of O157, O111, O26, O103 and O145 serogroups* | **YES** | **NO** |

|  |  |
| --- | --- |
| **2. If the answer to question 1 is NO please state deviations to ISO/TS 13136:2012** |  |

|  |  |
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| **3. For routine samples, briefly describe your enrichment process**  e.g. 25 g into 225 ml mTSB+N/BPW 41.5 °C for 24hr |  |

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| **4. Provide the details of your DNA extraction kit**  Include company name, kit name, protocol(s) if applicable:  e.g., Promega, Maxwell® 16 Cell DNA purification kit |  |

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| **5. If your DNA extraction process requires a platform please state platform used** |  |

**Laboratory Identification no. (Check) Lab No**

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| **6. State if you use conventional**  **RT PCR or real-time RT PCR** |  |

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| **7. If using a commercial kit for primer / probe / amplification mix**  Please provide company name, kit name: e.g., Bio-Rad, IQ-Check STEC SerO; Bio-Rad IQ-Check® STEC VirX |  |

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| **8. What volume of extracted DNA is used?** |  |

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| **9. Please complete the Cycle information tables** | **Pre-incubation:**   |  |  |  |  | | --- | --- | --- | --- | | **Time (hh:mm:ss)** |  | **Temperature (°C)** |  |   **Cycle information:**   |  |  |  |  |  |  |  | | --- | --- | --- | --- | --- | --- | --- | | **Parameter** | **Denaturisation** | **Cycling** | | | | **Cooling** | | **Cycles** | **x1** | **x**…….. | | | | **x1** | | **Temperature (°C)** |  | **Step 1:** | **Step 2:** | **Step 3:** | **Step 4:** |  | |  |  |  |  | | **Hold (hh:mm:ss)** |  |  |  |  |  |  |   **Other:** |

**Laboratory Identification no. Lab No**

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| **10. Amplification platform used**  e.g. Bio-Rad CFX96 Touch™ Deep Well RT-PCR Detection Systems |  |
| **11. Please use this space for any additional comments you feel are relevant or general comments about the scheme** |  |