Distribution of infectivity in animal tissue and body fluids

<u>Introduction</u>

- A.2.1 The following table (Table A2) presents a guide to the possible presence of TSE infectivity in various tissues and body fluids of cattle (exposed naturally or experimentally and orally to first passage BSE agent), sheep and goats (exposed naturally to scrapie agents and potentially to the BSE agent) irrespective of the stage of incubation. Table A2 has been updated in June 2010 taking account of the updated WHO Guidelines published in 2010 (WHO, 2010).
- A.2.2 This is the first time that the <u>WHO Guidelines</u> include tissues from Cervidae affected with Chronic Wasting Disease (CWD). CWD has not been reported in Europe despite some surveillance for it and there are no specific regulations in force. Should information on TSE infectivity in CWD be required, for example in regard to Cervidae in zoos or in animals in transit through our ports, please see updated 2010 WHO Guidelines.
- A.2.3 Throughout this document the term PrP^{TSE} is used to denote the disease-specific, partially protease resistant form of PrP. In the previous version the term PrP-res was used but for practical purposes PrP^{TSE} and PrP-res are the same.
- A.2.4 Additional notes are ascribed to certain entries to explain the differences (sometimes of interpretation) between the equivalent WHO table and Table A2, or to take account of new information. In case of doubt, please refer to the <a href="https://www.who.account.com/who.account.c
- A.2.5 Specified risk material (SRM) from cattle, sheep and goats in the UK and most other EU Member States, as at February 2007, is listed in a new table Table A3 in this document. Please refer to the <u>FSA</u> website for up-to-date information and definitions.
- A.2.6 The list of tissues in Table A2 is divided into three categories;
 - High infectivity tissues: Central nervous system (CNS) tissues and other tissues anatomically closely associated with them (in italics in the attached table). These become detectably infected in the later stages of all TSEs and into the clinical phase of disease.
 - Lower infectivity tissues: A range of peripheral tissues including some or all (depending on species) lymphoreticular system (LRS) tissues that may be infected from shortly after exposure.
 - Tissues that have either been bioassayed with no infectivity being detected, and/or tested for PrP^{TSE} and found negative.

A.2.7 A fourth category of tissues has not been tested in any of the species and therefore must remain as of uncertain infectivity. However, no tissue in this category is likely to have high infectivity but could become cross-contaminated in certain circumstances.

A.2.8 The following notes are of considerable importance when the table data are used in risk assessments:

- The precise relationship between the presence of PrP^{TSE} and infectivity is not certain. For example, the absence of detectable PrP^{TSE} does not necessarily mean the absence of infectivity. Conversely, detection of small amounts of PrP^{TSE} in a tissue does not necessarily mean that it would transmit disease in all circumstances.
- No account has been taken of the fact that a tissue having no detectable infectivity could become contaminated by inadequate care being taken at the point of collection. Such contamination could be direct (from other infected tissues) or indirect (from contaminated instruments). Furthermore, certain methods of stunning animals (such as those involving penetration of the cranium or use of a pithing rod) prior to killing by bleeding out may cause infected brain emboli to be dispersed into the blood stream and thence also to the heart, lungs and possibly to other organs and tissues.
- In the EU and some other countries certain tissues of ruminant species are categorised as SRM on the basis of a judgment that in an infected animal they would be, or might be infected. The list of tissues classed as SRM may be varied depending on the geographical BSE risk (GBR) ascribed to the host country. Some tissues in the current EC SRM list may not have demonstrated either infectivity or PrP^{TSE}, nevertheless any requirements of the law must be followed.
- In research laboratories, tissues from every challenged animal potentially present a risk. However, in diagnostic laboratories in the UK, tissues listed in Table A.2 and the WHO table will present a risk less frequently unless they come from a suspect case of TSE or from a high-risk group.

A.2.9 Extensive data on the distribution of infectivity, additional information and references can be found in the WHO guidelines at the WHO website:

http://www.who.int/bloodproducts/tse/WHO%20TSE%20Guidelines%20FINAL-22%20JuneupdatedNL.pdf. The WHO tables have copious notes which are not included in this ANNEX so the WHO tables should be consulted to aid interpretation of the indicated result, in any case of doubt or to extend knowledge.

The 2010 WHO update can be found at:

http://www.who.int/bloodproducts/tablestissueinfectivity.pdf

TABLE A2: Distribution of TSE infectivity in animal tissues and body fluids

	CATTLE		SHEEP AND GOATS	
TISSUE	INFECTIVITY	PrP ^{TSE}	INFECTIVITY	PrP ^{TSE}
HIGH INFECTIVITY	<u>′</u>			
Brain	+	+	+	+
Spinal ganglia				
(dorsal root ganglia) +	+	+	+
Spinal cord	+	+	+	+
Trigeminal ganglia	+	+	NT	+
Retina	+	NT	NT	+
Optic nerve	+	NT	NT	+
Pituitary gland	-	NT	+	+
LOWER INFECTIV	ITV			
Peripheral nervous				
Cauda equina	-	NT	NT	NT
Peripheral nerves	[+]	+	+	+
Autonomic ganglia	NT	+	NT	+
Lymphoreticular ti		·		·
Spleen	-	_	+	+
Lymph nodes	-	_	+	+
Nictitating Membrar	ne +	_	[+]	+
Thymus	-	NT	+	+
Tonsil	+	-	+	+
Alimentary tract			-	
Oesophagus	-	NT	[+]	+
Fore-stomachs	-	NT	[+]	+
Abomasum	-	NT	[+]	+
Duodenum	-	-	[+]	+
			Γ.1	•

Jejunum	-	+	[+]	+
lleum	+	+	+	+
Colon/caecum	-	-	+	+
Rectum	NT	NT	NT	+
Reproductive tissues				
Placenta	-	NT	+	+
Ovary	-	NT	-	-
Uterus	-	NT	-	-
Body fluids, secretions	and excretio	<u>ns</u>		
CSF	-	NT	+	-
Blood	-	?	+	?
Saliva	NT	NT	-	NT
Milk	-	-	+	[+]
Urine	-	NT	-	-
Faeces	-	NT	-	NT
Other tissues**				
Adrenal	[+]	+	+	-
Liver	-	NT	+	-
Pancreas	-	NT	+	NT
Nasal mucosa	-	NT	+	+
Blood vessels	-	NT	NT	+
Skeletal Muscle*	[+]	NT	[+]	+
Salivary gland	-	NT	+	NT
Mammary gland/udder	-	NT	-	+
Skin	-	NT	-	+
Adipose tissue	-	NT	NT	NT
Heart/pericardium	-	NT	-	NT
Lung	-	NT	-	-
Kidney	-	-	[+]	+
Bone marrow	(+)	NT	+	NT
Tongue	-	NT	[+]	+

NO DETECTABLE INFECTIVITY

Reproductive tissues

Testis	-	NT	-	-
Prostate/Epididymis/				
Seminal vesicle	-	NT	-	-
Semen	-	NT	-	-
Placenta fluids	-	NT	NT	NT
Fetus	-	NT	-	-
Embryos	-	NT	?	NT
Musculo-skeletal tissues	<u>s</u>			
Bone	-	NT	NT	NT
Tendon	-	NT	NT	NT
Other tissues**				
Trachea	-	NT	NT	NT
Thyroid gland	NT	NT	-	NT
Body fluids, secretions	and excretion	าร**		
Colostrum	(-)	-	(?)	NT
Cord blood	-	NT	NT	NT

Key:

Italic type	Tissues included due to close anatomical proximity to high infectivity tissues
+	Positive transmission or presence of PrP ^{TSE} detected
-	Bioassay negative or PrPTSE not detected
NT	Bioassay not undertaken or PrP ^{TSE} not tested for
NA	Not applicable
No detectable infectivity	Tissues not included in other parts of the table but
•	bioassayed and /or tested for PrPTSE with negative
	result.
()	Limited or preliminary data
?	Uncertain interpretation
	Infectivity or PrP ^{TSE} data based exclusively on
-	bioassays in transgenic (Tg)mouse over-expressing the
	bioassays in transgenic (Tg)mouse over-expressing the PrP-encoding gene, or PrP ^{TSE} amplification methods

^{*} In cattle, preliminary data indicate a low level of infectivity in the *M. semitendinosus* from one cow with clinical BSE. A homogenate of this muscle was inoculated i/c-i/p into ten highly BSE-sensitive transgenic mice and PrP^{TSE} was detected in one mouse (Buschmann and Groschup, 2005). No transmissions have been reported when various bovine tissues have been inoculated into either susceptible mice or cattle by the i/c-i/p or i/c routes respectively. Concern has been expressed about possible infectivity in lingual tonsillar

tissue at the root of the tongue, though the tongue itself shows no detectable infectivity (Wells *et al.*, 2005).

** Some tissues tested in humans have not been tested in animals such as cornea, some dental tissues, sweat, tears, mucus and bile.

SPECIFIED RISK MATERIAL

A.2.10 Specified Risk material (SRM) includes those tissues of cattle, sheep and goats which are known to, or might potentially, harbour detectable BSE infectivity in infected animals. SRM is excluded from the human food and animal feed chains and cannot be used for any other purpose. The tissues which fall within the current definition of SRM are listed on the UK Food Standards Agency website http://www.food.gov.uk/safereating/animaldiseases/bse/what/beef/controls and are given below in Table A3.

TABLE A3 Specified risk material in the EU (as at November 2006)

Specified Risk Mat	Specified Risk Material in all Member States from 25 May 2006		
Cattle	All ages		
	The tonsils, the intestines, from the duodenum to the rectum, and the mesentery		
	Over 12 months		
	 Skull excluding the mandible but including the brains and eyes, and spinal cord 		
	Over 24 months		
	 Vertebral column, excluding the vertebrae of the tail the spinous and transverse processes of the cervical, thoracic and lumbar 		
	vertebrae, the median sacral crest and the wings of the sacrum, but including the dorsal root ganglia		
Sheep and goats	All ages		
	 The spleen and the ileum 		
	Over 12 months (or permanent incisor erupted)		
	 Skull including the brains and eyes, tonsils, spinal cord 		

REFERENCES

A comprehensive list of references is given in the WHO Guidelines which should be consulted when further detail is required. The following are selected, relatively recent, publications of relevance to the Tables above.

General

WHO. Guidelines on tissue infectivity distribution in transmissible spongiform encephalopathies; June 2006.

http://www.who.int/bloodproducts/tse/WHO%20TSE%20Guidelines%20FINAL-22%20JuneupdatedNL.pdf

WHO. Guidelines on tissue infectivity distribution in transmissible spongiform encephalopathies; Updated 2010.

Bovine Spongiform Encephalopathy

Buschmann A, Groschup MH. Highly BSE sensitive transgenic mice confirm essential restriction of infectivity to the nervous system in clinically diseased cattle. *J. Infect Dis* 2005; **192**:934-42

Everest SJ, Thorne LT, Hawthorn JA, Jenkins R, Hammersley C, Ramsay AM, Hawkins SA, Venables L, Flynn L, Sayers R, Kilpatrick J, Sach A, Hope J, Jackman R. No abnormal prion protein detected in the milk of cattle infected with the bovine spongiform encephalopathy agent. *J Gen Virol* 2006; **87**: 2433-2441

Hoffmann C, Ziegler U, Buschmann A, Weber A, Kupfer L, Oelschlegel A, Hammerschmidtand B, Groschup MH. Prions spread via the autonomic nervous system from the gut to the central nervous system in cattle incubating bovine spongiform encephalopathy. *J Gen Virol* 2007; **88**: 1048–55

Iwamaru Y, Okubo Y, Ikeda T, Hayashi H, Imamura M, Yokoyama T, Shinagawa M.

PrPSc distribution of a natural case of bovine spongiform encephalopathy. In: Kitamoto T, ed. *Prions. Food and Drug Safety.* Springer Verlag, New York, 2005

Wells GAH, Spiropoulos J, Hawkins SAC, Ryder SJ. Pathogenesis of experimental bovine spongiform encephalopathy (BSE): pre-clinical infectivity in tonsil and observations on lingual tonsil in slaughtered cattle. *Vet Rec* 2005; **156**: 401-7.

Scrapie

Andreoletti O, Lacroux C, Chabert A, Monnereau L, Tabouret G, Lantier F, Berthon P, Eyenne F, Lafond-Benestad S, Elsen J-M, Schelcher F. PrP(Sc) accumulation in placentas of ewes exposed to natural scrapie: influence of foetal *PrP* genotype and effect on ewe-to-lamb transmission. *J Gen Virol* 2002; **83**: 2607-16.

Casalone C, Corona C, Crescio MI, Martucci F, Mazza M, Ru G, Bozzetta E, Acutis PL, Caramelli M. Pathological prion protein in the tongues of sheep infected with naturally occurring scrapie. *J Virol* 2005; **79**: 5847-9.

Specified Risk Material

FSA (Food Standards Agency UK). List of tissues which fall within the current definition of Specified Risk Material. 2006:

http://www.food.gov.uk/safereating/animaldiseases/bse/what/beef/controls